

BI/CH 422/622

OUTLINE:

Introduction and review
Transport
Glycogenolysis
Glycolysis

Introduction & overview; 2 phases
Phase I
Phase II
Summary: logic, energetics, labeling studies

Other sugars

Pasteur: Anaerobic vs Aerobic

Fermentations: anaerobic fates of pyruvate

Lactate-lactate dehydrogenase

Exam-1 material Acetoacetate decarboxylase

Exam-2 material Ethanol-pyruvate decarboxylase & alcohol dehydrogenase

Pyruvate oxidation:
aerobic fates of pyruvate
pyruvate dehydrogenase complex

Krebs' Cycle

How did he figure it out?

Overview

8 Steps

Citrate Synthase

Aconitase

Isocitrate dehydrogenase

Ketoglutarate dehydrogenase

Succinyl-CoA synthetase

Succinate dehydrogenase

Fumarase

Malate dehydrogenase

Energetics; Regulation

Summary

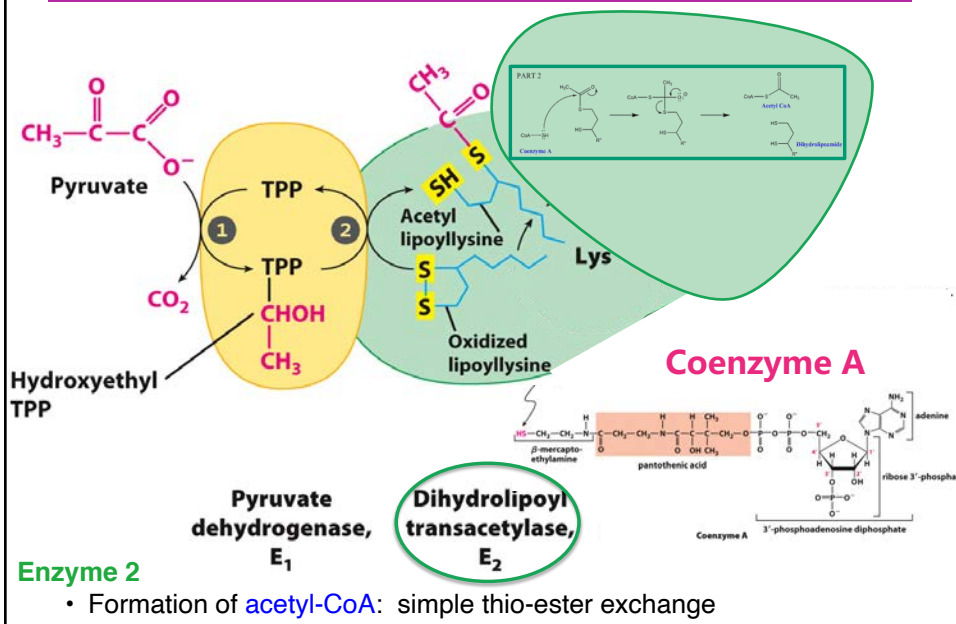
Oxidative Phosphorylation

Electron Transport

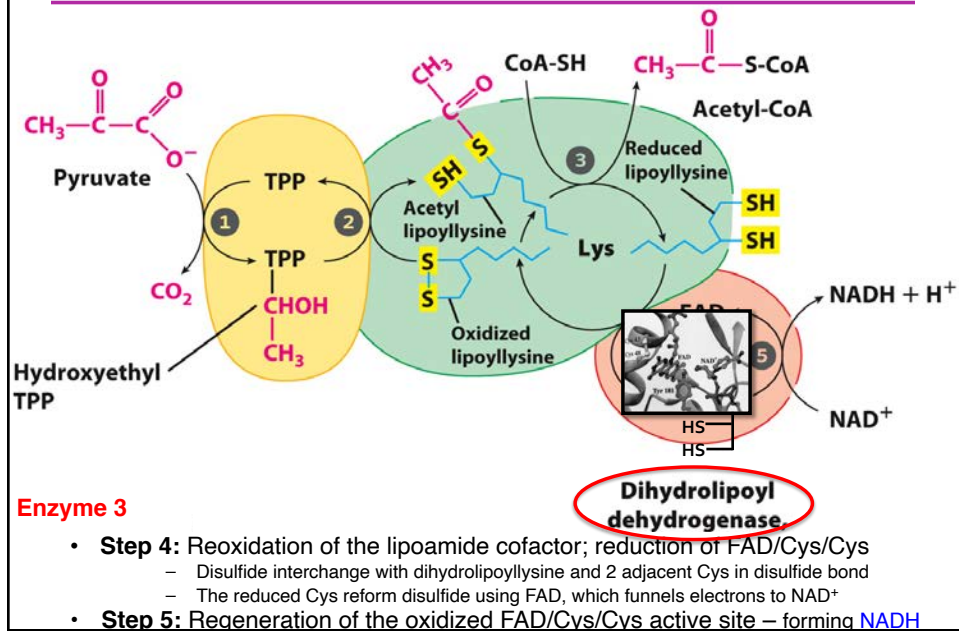
Chemiosmotic theory

ATP synthesis

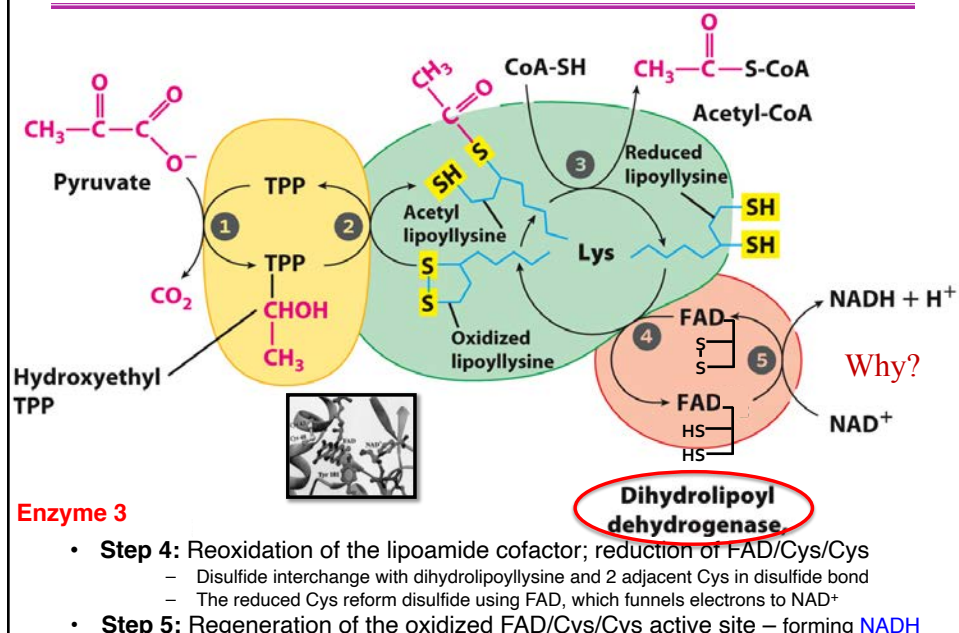
Pyruvate Oxidation



Pyruvate Oxidation



Pyruvate Oxidation



Pyruvate Oxidation

Enzyme 3: Dihydrolipoamide dehydrogenase

Chemical structures of FAD/FADH2 and FMN/FMNH2:

Oxidized form (yellow): $\lambda_{max} = 450 \text{ nm}$

Reduced form (colorless): $\lambda_{max} = 490 \text{ nm}$

Semiquinone form (blue): $\lambda_{max} = 570 \text{ nm}$

Semiquinone anion (red): $\lambda_{max} = 600 \text{ nm}$

Redox Potentials:

$$\Delta E^{\circ} = E^{\circ}_{(reduction)} - E^{\circ}_{(oxidation)}$$

$$\Delta E^{\circ} = E^{\circ}_{(NAD^{+})} - E^{\circ}_{(FAD)}$$

$$= -0.320 \text{ V} - (+0.031 \text{ V}^*)$$

$$= -0.351 \text{ V}$$

$$\Delta G^{\circ} = -n \mathcal{F} \Delta E^{\circ}$$

$$= -(2)(23.06 \text{ kcal/V} \cdot \text{mol}^{-1})(-0.351 \text{ V})$$

$$= +16 \text{ kcal mol}^{-1}$$

*from Maeda-Yorita et al., (1994) Biochem. 33, 6213

Pyruvate Oxidation

Enzyme 3: Dihydrolipoamide dehydrogenase

Standard Redox Potentials E°

Half reaction	$E^{\circ}(\text{V})$
Succinate + $\text{CO}_2 + 2\text{H}^+ + 2e^- \rightleftharpoons \alpha\text{-ketoglutarate} + \text{H}_2\text{O}$	-0.670
Acetate + $2\text{H}^+ + 2e^- \rightleftharpoons \text{acetaldehyde}$	-0.581
$2\text{H}^+ + 2e^- \rightleftharpoons \text{H}_2$	-0.421
$\alpha\text{-ketoglutarate} + \text{CO}_2 + 2\text{H}^+ + 2e^- \rightleftharpoons \text{citrate}$	-0.380
Cysteine + $2\text{H}^+ + 2e^- \rightleftharpoons 2 \text{ cysteine}$	-0.340
$\text{NAD}^+ + 2\text{H}^+ + 2e^- \rightleftharpoons \text{NADH} + \text{H}^+$	-0.320
$\text{NADP}^+ + 2\text{H}^+ + 2e^- \rightleftharpoons \text{NADPH} + \text{H}^+$	-0.324
Acetaldehyde + $2\text{H}^+ + 2e^- \rightleftharpoons \text{ethanol}$	-0.197
Pyruvate + $2\text{H}^+ + 2e^- \rightleftharpoons \text{lactate}$	-0.185
Oxaloacetate + $2\text{H}^+ + 2e^- \rightleftharpoons \text{malate}$	-0.166
$\text{FAD} + 2\text{H}^+ + 2e^- \rightleftharpoons \text{FADH}_2$	0.031
Fumarate + $2\text{H}^+ + 2e^- \rightleftharpoons \text{succinate}$	0.031
Ubiquinone + $2\text{H}^+ + 2e^- \rightleftharpoons \text{ubiquinol}$	0.045
2 cytochrome $b_{558} + 2e^- \rightleftharpoons 2 \text{ cytochrome } b_{558}$	0.070
2 cytochrome $c_{550} + 2e^- \rightleftharpoons 2 \text{ cytochrome } c_{550}$	0.254
2 cytochrome $a_{350} + 2e^- \rightleftharpoons 2 \text{ cytochrome } a_{350}$	0.385
$1/2 \text{ O}_2 + 2\text{H}^+ + 2e^- \rightleftharpoons \text{H}_2\text{O}$	0.816

Redox Potentials:

$$\Delta E^{\circ} = E^{\circ}_{(reduction)} - E^{\circ}_{(oxidation)}$$

$$\Delta E^{\circ} = E^{\circ}_{(NAD^{+})} - E^{\circ}_{(FAD)}$$

$$= -0.320 \text{ V} - (+0.031 \text{ V}^*)$$

$$= -0.351 \text{ V}$$

$$\Delta G^{\circ} = -n \mathcal{F} \Delta E^{\circ}$$

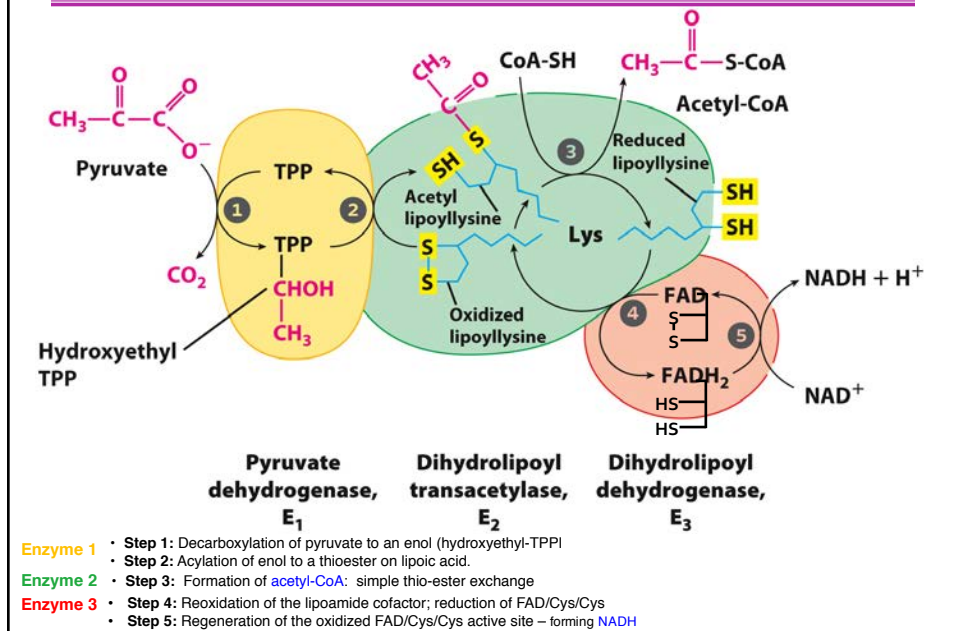
$$= -(2)(23.06 \text{ kcal/V} \cdot \text{mol}^{-1})(-0.351 \text{ V})$$

$$= +16 \text{ kcal mol}^{-1}$$

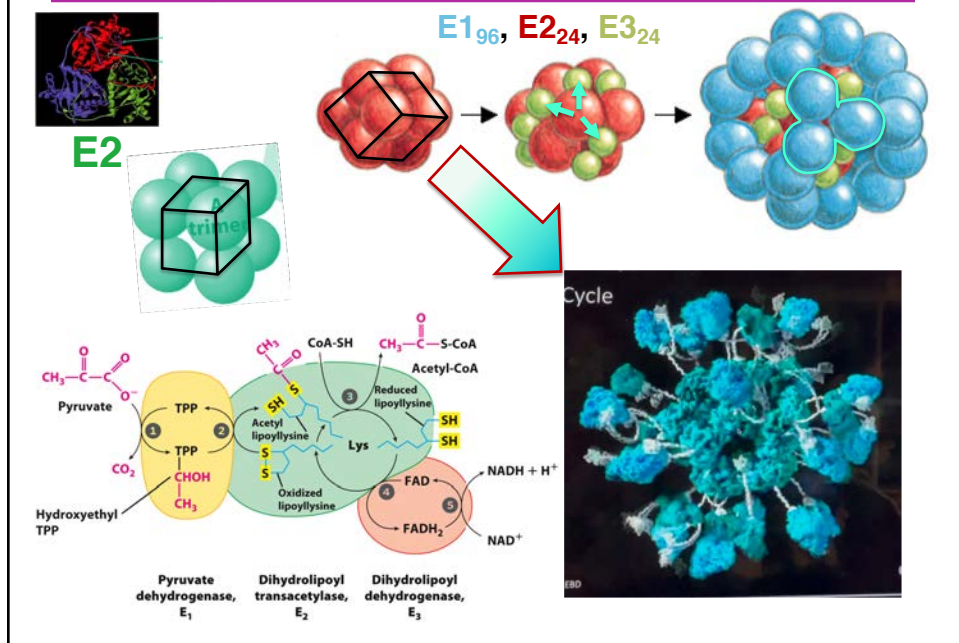
If use S-S, with $E^{\circ}_{(S-S)} = -0.34 \text{ V}$, $\Delta G^{\circ} = -0.92 \text{ kcal/mol}$

*from Maeda-Yorita et al., (1994) Biochem. 33, 6213

Pyruvate Oxidation



Pyruvate Oxidation



Pyruvate Oxidation

Overall Reaction of PDC

Pyruvate + Coenzyme-A (CoASH) + NAD⁺

PDC (TPP, lipoic acid, FAD) $\Delta G^{\circ} = -8 \text{ kcal/mol}$

$\text{CO}_2 + \text{Acetyl-Coenzyme-A (Ac-CoA)} + \text{NADH} + \text{H}^+$

Pyruvate Oxidation

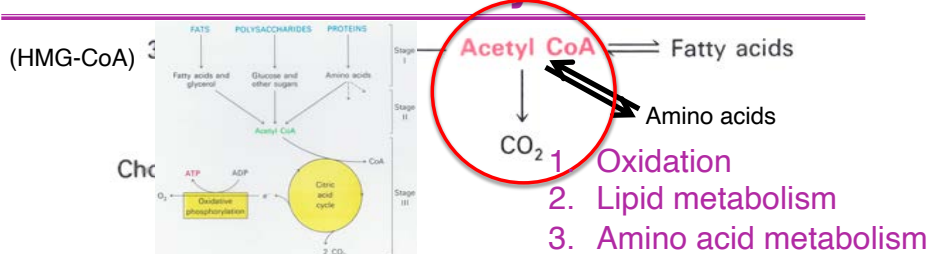
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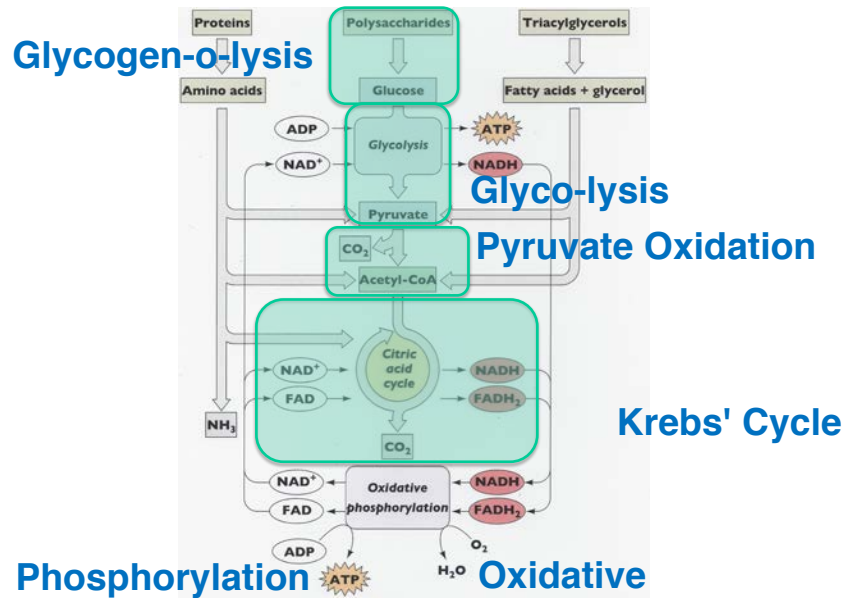
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Fates of Acetyl CoA



The Citric Acid Cycle



The Citric Acid Cycle

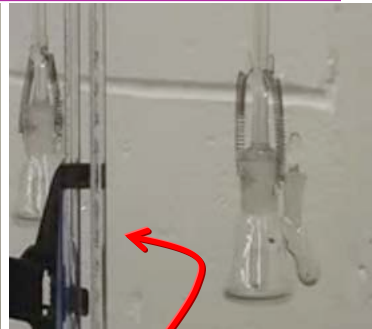
a.k.a. Krebs Cycle,
a.k.a. Tricarboxylic Acid Cycle (TCA)



Time B.C. (Before the Cycle)



Otto Warburg
1883-1970



Manometer

Warburg Apparatus

-respiration

-Measure rates of O₂ consumption

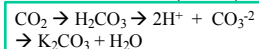
UTube instructions

(<http://youtu.be/M-HYbZwN43o>)

Substrates
(e.g., glucose)

Tissues

CO₂ trap
(KOH)

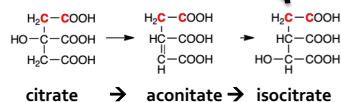
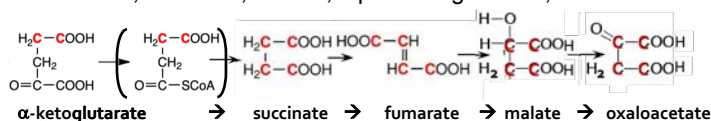


Time B.C. (Before the Cycle)

In 1920 BC, what was known about respiration?

- 1) Glycolysis gives rise to pyruvate
- 2) Adding 1 mole pyruvate to respiring tissues in a Warburg apparatus, there are 2.5 moles O₂ consumed:

$$2^{1/2}\text{O}_2 + \text{C}_3\text{H}_4\text{O}_3 \rightarrow \rightarrow \rightarrow \rightarrow 3\text{CO}_2 + 2\text{H}_2\text{O}$$
- 3) Any intermediate in the process will be oxidized at a rate \geq pyruvate
- 4) Many intermediates were tried, but few met this criteria, they were: succinate, fumarate, malate, alpha-ketoglutarate, etc.



Albert Szent-Györgyi
1893-1986

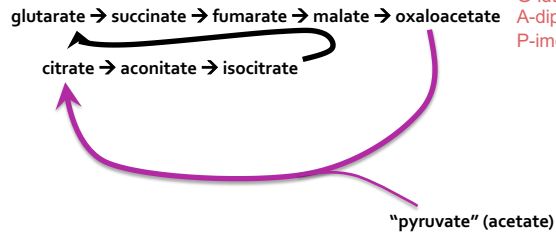
5) Others had already worked out several compounds and their interconversion. Specifically, Albert Szent-Györgyi had worked out the interconversion of the dicarboxylic acids. Carl Martinus worked out the interconversion of the tricarboxylic acids

6) In 1937, with help of German biochemist Franz Koop, Carl Martinus, demonstrated a series of reactions using citrate that produced α -ketoglutarate. Thus tricarboxylic acid and dicarboxylic acids would be interconverted with loss of CO₂, but also support respiration.

Time B.C. (Before the Cycle)



Hans Krebs
1900-1981

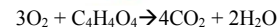
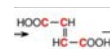


O-leic
M-alonic
S-uccinic
G-lutaric
A-dipic
P-imelic

Krebs confirmed that the pathway was consistent with succinate, fumarate, and malate proved to be useful because all these molecules increased oxygen consumption in the pigeon breast muscle.

He observed much more than 3:1!!

The first clue came from an experiment with fumarate. Krebs did careful measurements using the Warburg manometer. Fumarate gave greater than expected oxygen consumption in the pigeon breast muscle.



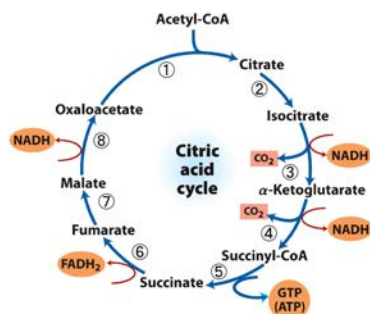
$\therefore 1 \mu\text{mole fumarate would consume } 3 \mu\text{mole O}_2$

- 1) Malonic acid inhibition of the **succinate → fumarate** step prevented this increase...BUT, succinate **accumulated**!
- 2) How can fumarate give rise to succinate? There must be a cycle
- 3) Tested by showing that using succinate or fumarate you could detect the formation of citrate.

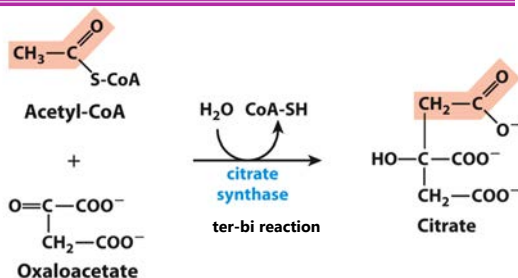
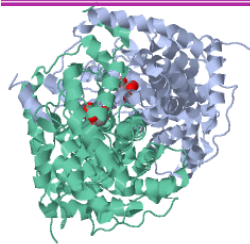
Later in 1937, he proposed that pyruvate would combine with oxaloacetate to make citrate in a cycle he called the Citric Acid Cycle. Later, Fritz Lipmann showed that it was acetyl-CoA and not pyruvate.

The Citric Acid Cycle

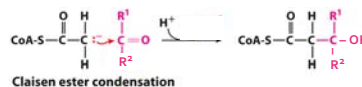
- Step 1: C-C bond formation between acetate (2C) and oxaloacetate (4C) to make citrate (6C)
- Step 2: Isomerization via dehydration/rehydration
- Steps 3–4: Oxidative decarboxylations to give 2 NADH
- Step 5: Substrate-level phosphorylation to give GTP
- Step 6: Dehydrogenation to give FADH₂
- Step 7: Hydration
- Step 8: Dehydrogenation to give NADH



The Citric Acid Cycle: Citrate Synthase



- Joining of acetyl-CoA and oxaloacetate with **C-C bond formation**
- Highly thermodynamically **favorable/irreversible** ($\Delta G^\circ = -7.7$ kcal/mol)
 - regulated by substrate availability and product inhibition
- Activity largely depends on [oxaloacetate].
- Rate-limiting step of CAC
- **Uses acid/base catalysis**
 - Carbonyl of oxaloacetate is a **good electrophile**.
 - Methyl of acetyl-CoA is **not** a good nucleophile...
 - ...unless activated by deprotonation to form a **carbanion**.



The Citric Acid Cycle: Citrate Synthase

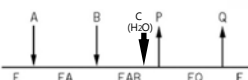
Mechanism

- Conformational change occurs upon binding oxaloacetate.
- Avoids unnecessary hydrolysis of thioester in acetyl-CoA

a) Open conformation:

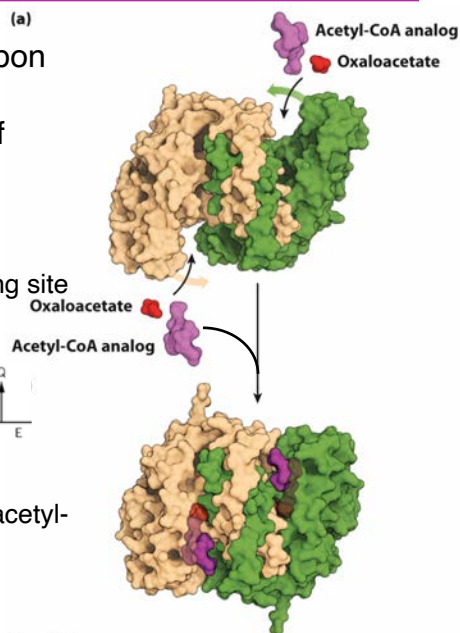
Free enzyme does not have a binding site for acetyl-CoA. Ordered binding.

Sequential ordered ter bi



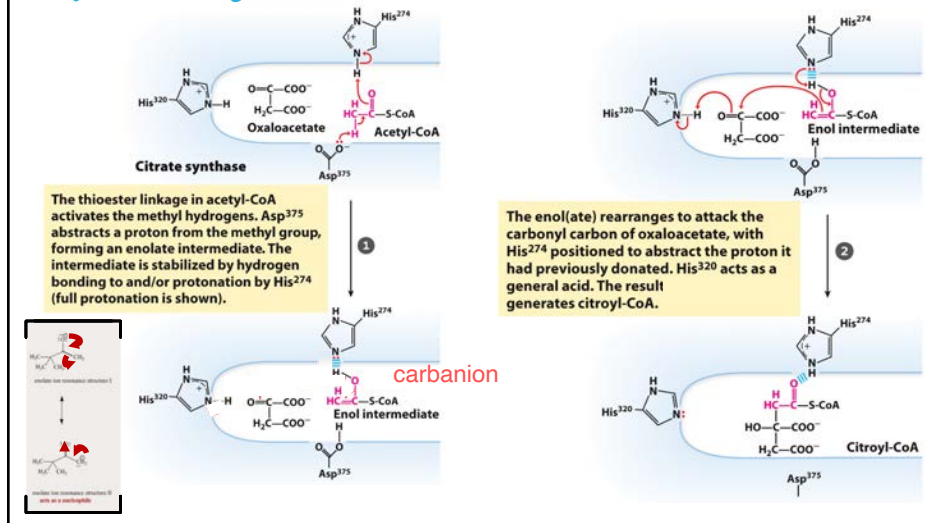
b) Closed conformation:

Binding of OAA creates binding for acetyl-CoA.
Reactive carbanion is protected.



The Citric Acid Cycle: Citrate Synthase

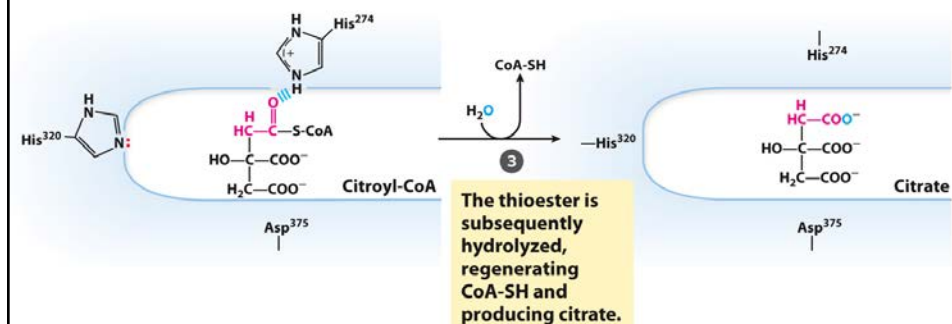
Mechanism



The Citric Acid Cycle: Citrate Synthase

Mechanism

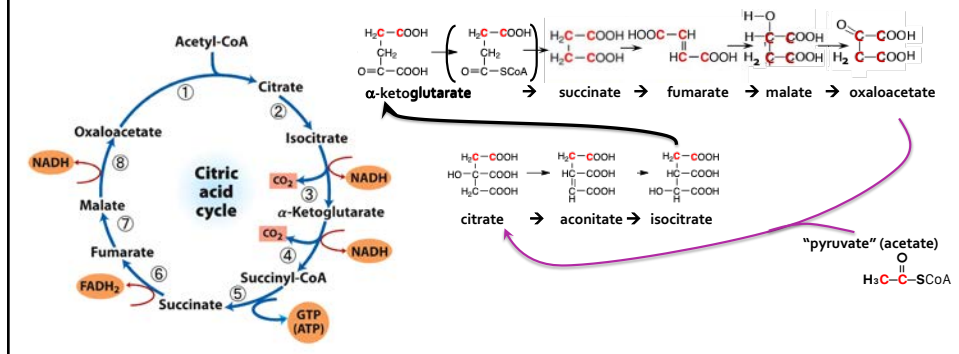
Hydrolysis of Thioester; citryl-CoA



The Citric Acid Cycle

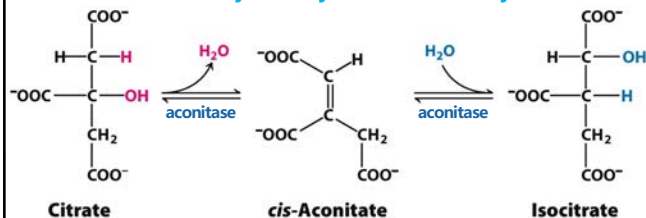
Citrate Synthase

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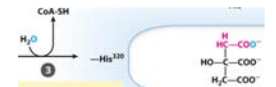


The Citric Acid Cycle: Aconitase

Isomerization by Dehydration/Rehydration



- Elimination of H₂O from the symmetrical molecule, citrate, gives a cis C=C bond.
 - lyase



Rationale:

- Citrate, a tertiary alcohol, is a poor substrate for oxidation.
- Isocitrate, a secondary alcohol, is a good substrate for oxidation.

Thermodynamically unfavorable/reversible ($\Delta G^\circ = +3.2$ kcal/mol)

- product concentration kept low to pull forward; citrate tends to "pool" with higher conc.

Dehydration & Addition of H₂O to cis-aconitate is stereospecific.

- This was initially very confusing to bio/organic chemists
- Only R-isocitrate is produced by aconitase.
- A biochemist names A.G. Ogston clarified the situation by realizing that the enzyme spatially templates this symmetrical molecule by binding in only one way (e.g., clockwise or counter clockwise, not both)
- Distinguished by three-point attachment to the active site

Realize that biochemists could distinguish the two acetates by running the citrate synthase reaction with H₂¹⁸O